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COMPILED BY

ALICE G. C. GRANDISON, B.Sc.

PAULINE CURDS, B.Sc.

and

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ARTICLE

OF THE
CONSTITUTION
OF THE UNITED STATES
OF AMERICA

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Behaviour.—Role of the pineal system in amphibian behaviour, Adkins & Zweifel; Importance of habits in evolutionary studies of spadefoot toads, Bragg (1); Aggregational behaviour of spadefoot tadpoles, Bragg, (2); Summer surface activity in California salamanders, Brame, A. (1); Differential behaviour of *Xenopus laevis*, Haurich (1); Hierarchical behaviour in *Xenopus laevis*, Haurich (2); Activity and behaviour of *Rana sylvatica*, Heatwole (1); Habits of amphibians in captivity and in natural biotopes modified by man, Heusser; Change in daily activity of *Rana ridibunda* in relation to temperature, Pashkova & Sudarev; Habits of *Megalobatrachus japonicus davidianus* in aquarium, Šafránek;

Gregarious behaviour of *Rhinophrynus dorsalis* tadpoles, **Stuart**; Activities of frogs in terrarium, **Südkow**; Habits of *Hyperolius h. horstokii*, **Visser**; *Rana clamitans* and *R. pipiens* apparently more active in rainy weather on basis of number caught in mousetraps, **Whitaker**.

Breeding and Parental Habits.—Mating behaviour of *Euproctus asper* in captivity, **Ahrenfeldt**; Breeding site and habits of a spadefoot, toad and frog breeding congress, **Bragg** (2); Observations on ♀ *Desmognathus fuscus* with eggs in a relatively dry habitat, **Brode**; Mating habits of *Hymenochirus boulengeri*, **Haas**; *Necturus* breeding habits, **Harris, J. P., Jr.**; Part played by environmental factors in amphibian reproductive activity, **Heusser**; Mating season of *Rana temporaria* in Poland, **Juszczyk**; Calls of breeding congresses of some Jigalong (Australia) species; description of sites, **Lindgren & Main**; *Hyla faber* breeding habits, **Lutz**; Parental care in anurans, **Lynn**; Role of breeding site in anuran reproductive isolation, **Mecham** (3); Breeding habits of New South Wales frogs, **Moore, J. A.** (2); Egg mass and attending ♀ *Desmognathus wrighti*, **Organ** (1); Breeding behaviour and brood care of *Pipa pipa*, **Rabb**; Breeding habits of *Hyla crucifer* and *H. c. bartramiana*, **Roth**; Breeding behaviour of *Rana temporaria*, **Savage, R. M.**; *Bufo exilis* and *B. nelsoni* breeding habits, **Savage & Schnierer**; Egg laying habits of *Cophixalus* and *Asterophrys wilhelmani* in New Guinea, **Tyler** (2); Breeding sites of some Anura, **Van Dijk**; *Hymenochirus boulengeri* breeding habits, **Yuhua**.

Burrowing Habits.—Burrowing habits of desert amphibians, **Chew**.

Cannibalism.—Records of cannibalism in *Plethodon cinereus*, **Heatwole & Test**; Absence of cannibalism in *Rhinophrynus dorsalis* tadpoles from Tikal, Guatemala, **Stuart**.

Courtship.—Courtship behaviour of *Ambystoma macrodactylum croceum*, **Anderson, J. D.** (1); Courting behaviour of *Triturus*, **Gauss**; Courtship behaviour of *Desmognathus wrighti*, **Organ** (1); Courtship behaviour and season in *Desmognathus*, **Organ** (2); Local variation of mating behaviour in *Triturus pyrrhogaster*, **Sawada**.

Diseases.—Amphibian diseases with notes on their therapeutics, **Reichenbach-Klinke**.

Distribution.—Ranges of allopatric and sympatric populations of *Leptodactylus ocellatus* and *L. chaguenensis* in Argentina, **Cei & Bertini** (2); Decreasing frequency of *Bombina bombina* in Sweden, **Curry-Lindahl** (1); South American distribution of Argentine Brachycephalidae, **Gallardo** (7); Local distribution and habitat selection by *Rana sylvatica*, **Heatwole** (1); Correlation between critical thermal maxima, habitat and range of salamanders, **Hutchinson**; *Bombina bombina* and *B. variegata* in Czechoslovakia, **Lác** (1); Altitude limits of amphibians in Orava River basin, Czechoslovakia, **Lác** (2); Distributional ranges and locality records in Canada and Alaska, **Logier & Toner**; Geographic distribution of *Rana tigrina*, **Manacas**; Distributional ranges of *Bombina* in Krakow-Chrzanów and R. Wisla regions, Poland, **Michalowski** (1); Patterns of distribution of

Australian frogs, **Moore, J. A.** (2); New Central American records for *Rhinophrynus dorsalis*, **Nelson & Hoyt**; Local distribution of *Desmognathus* in Virginia, **Organ** (2); Distribution patterns in southern Africa, **Poynton** (1); Illinois amphibians, a critical review of species and their distribution, **Smith, P. W.**; Home range and population notes for *Plethodon cinereus*, **Taub**; Colour variant of *Plethodon cinereus* associated with glacial boundaries, **Thurrow**.

Ecological Associations.—Association of *Rana tigrina* with buffalo in Portuguese India, **Manacas**; Of *Desmognathus* in Virginia, **Organ** (2); Amphibia of swamps and marshes in S.W. Spain, **Valverde**.

Economics.—Method of eating frogs in New Guinea, **Tyler** (2).

Effects of Environment.—Relation of temperature and rainfall to calling and spawning activity in a mixed anuran population, **Blair, W. F.** (2); Effect of the volcano Parícutin on vertebrates, **Burt**; Effect of pond and laboratory rearing on cell division in *Ambystoma* larvae, **Chiakulas & Scheving**; Seasonal and lunar variation in numbers of mating toads in Bandung, Java, **Church** (2); Autumnal pause in morphogenesis of *Alytes* tadpoles, **Discol & Bounhiol**; Disappearance of *Salamandra salamandra* from Limburg, Holland, **Fonteyne** (1); Disappearance of *Bombina variegata* from Limburg, Holland, **Fonteyne** (2); Importance of environment in behaviour, **Heusser**; Seasonal and altitudinal variation in diet in Congolese amphibians, **Inger & Marx**; Influence of temperature on life and activity of *Rana temporaria*, **Juszczyk**; Water levels as an environmental factor in *Bufo boreas* breeding season, **Metter**; Effects of temperature on *Rana ridibunda* under natural conditions, **Pashkova & Sudarev**; Growth inhibitory substances produced by crowded *Rana pipiens* tadpoles, **Rose, S. M. & F. C.**; Influence of external factors on spawning date of the common frog, **Savage, R. M.**; Predictable reactions of a *Plethodon* population toward environmental conditions, **Taub**; Seasonal variation in feeding habits of *Rana ridibunda* in Roumania, **Vancea, Mindou & Simionescu**.

Enemies and Defence.—Predation by caddisfly larvae on *Ambystoma maculatum* eggs, **Murphy**; Parasites and predators of juvenile *Rana temporaria*, **Savage, R. M.**; *Cophixalus parkeri* eaten by *Halcyon sancta*, **Tyler** (1).

Feeding Habits.—Stomach contents of *Siren intermedia* from North Carolina, **Collette & Gehlbach**; *Plethodon jordani* found eaten by *Desmognathus quadramaculatus*, **Huhey & Brandon**; Dominant foods, seasonal and altitudinal variation in diet, and species—prey size relations in Congolese Amphibia, **Inger & Marx**; Arthropods and nematodes in *Herpele squalostoma* stomach. Vegetable matter in *Geotrypetes* stomach, **Laurent** (2); Stomach contents of *Rana tigrina* from Portuguese India, **Manacas**; Effect of diet on tadpole growth: experiments using snails, **Mihail & Asandei**; Feeding habits of adult *Taricha granulosa* and *T. rivularis*, **Facker**; Stomach analyses of *Rana ridibunda* in Roumania, **Vancea, Mindou & Simionescu**; Stomach contents of mouse-trapped *Rana pipiens* and *R. clamitans* from Ithaca, N.Y., **Whitaker**.

Hibernation.—Hibernation of *Bufo hemiophys*, **Breckenridge & Tester**; Hibernation of *Bufo* by internal reactions, independent of temperature, **Heusser**; Hibernation of *Rana temporaria* in Poland, **Juszczyk**; Hibernation survival studies on *Plethodon cinereus*, **Taub**.

Homing.—Homing experiments on *Aneides* experimentally removed from habitat, **Gordon, R. E.** (2); Homing migrations of *Rana esculenta* recorded in Germany, **Sanden-Guja**; Homing behaviour in *Taricha rivularis*, **Twitty** (1).

Larval Stages.—Ecology of *Rana temporaria* tadpoles and their anatomy and physiology, **Savage, R. M.**

Life History.—Life history of *Ambystoma rosaceum*, **Anderson, J. D.** (2); Of *Rana temporaria* and *Triturus helveticus* in high mountain lakes in S. France, **Angelier**; Seasonal and lunar changes in reproductive cycle of *Bufo melanostictus* from Bandung (Java), **Church** (2); Of *Necturus*, **Harris, J. P., Jr.**; Types of life history in Amphibia, **Lynn**; Breeding season of *Bufo boreas* affected by water levels, **Metter**; Life history of *Desmognathus* in Virginia, **Organ** (2).

Locomotion.—Stances and walking movements of New Zealand frogs, **Barwick**; Bipedalism in frogs, **Dubrul**; Biomechanics of *Rana catesbeiana* jumping at prey, **Gans** (4); Locomotive activity of sympatric salamanders in the Appalachians, **Gordon, R. E.** (1); Importance of pelvic girdle in locomotion, **Whiting**.

Moult.—Hormonal control of moulting in toads, **Jørgensen & Larsen**.

Movement and Migration.—Local movements of *Bufo hemiophys* in Minnesota, studied by marking with Ta¹⁸², **Breckenridge & Tester**; Migrations of *Taricha granulatus*, **Carl**; Toad migrations in Camberley, **Davison**; Seasonal migrations of *Rana temporaria* in Poland, **Juszczyk**; Seasonal movements of *Bufo bufo*, **Moore, H. J.**; Breeding migration by *Taricha rivularis*, **Twitty** (1); Natural dispersal of *Aneides aeneus* recorded in Kentucky, **Williams & Gordon**.

Nesting Habits.—Of *Necturus*, **Harris, J. P., Jr.**; Functions of brooding behaviour in ♀ *Plethodon cinereus*, **Highton & Savage**; Nest-building by ♂ *Hyla faber*, **Lutz**; Nesting site of *Desmognathus wrighti*, **Organ** (1); Nesting sites, egg laying and egg masses of *Desmognathus*, **Organ** (2); Nesting site and egg masses of *Pseudotriton porphyriticus*, **Organ** (3); Foam nests of *Rhacophorus leucomyx*, **Stolk** (10).

Parasitism.—Intestinal Protozoa of *Rana septentrionalis*, **Camara & Euttre**; Plerocercosis of *Bufo arenarum*, **Funes & Padilla**; *Rana catesbeiana* host to a new tapeworm, **Jones, Cheng & Gillespie**; Haematocysts of Australian frogs, **Mackerras, M. & I. M.**; Amphibian parasitic diseases, **Reichenbach-Klinke**; *Brachycolium salamandrae* in the small intestine of *Salamandra salamandra*, **Szabó**; *Rana esculenta* as second intermediate host in cycle of *Ascoctyle branchialis* sp. nov. (Trematoda), **Timon-David**; List of Amphibia as hosts of nematode parasites, **Walton** (1); Amphibia as hosts of cestodes and acanthocephalans, **Walton** (2).

Population Studies.—Frequency of mutations in X-irradiated populations of *Bufo valliceps*, **Blair, W. F.** (1); Seasonal patterns of reproductive activity in mixed anuran population and importance in respect to possible interpopulation competition, **Blair, W. F.** (2); Distribution patterns and population densities in the submontane and montane zones of Cuernos de Negros, Philippine Islands, **Brown & Alcalá**; Electrophoretic patterns of two sympatric and allopatric Argentine populations of *Leptodactylus ocellatus* and *L. chaquensis*, **Cei & Bertini** (2); Populations of *Desmognathus fuscus* from Louisiana and Arkansas, compared, **Chaney**; Population dynamics, **Edelstein**; Amphibia in R. Elbe lowlands, Germany, **Garms**; *Eupsophus* now known from Chile (lat. 32° S. to 53° S.), represented by 5 species sympatric over a large area within the Valdivian Forest faunistic zone, **Grandison** (1); *Crinia insignifera* populations on Rottnest Island, Australia; inheritance, fecundity, viability, yearly changes in morph frequency, and population size analyses, **Main**; Population density studies on anuran tadpoles, **Muzilek**; Local distribution, life history and population dynamics of *Desmognathus* in Virginia, **Organ** (2); Colour variations in *Acris crepitans* populations, **Plyburn** (1); Population studies on *Taricha* in Sonoma Co., **Twitty** (1).

Technique.—Use of arthropods to prepare osteological specimens, **Banta** (1); Technique for collecting in urban areas, **McCoy**; Collecting methods in a population study of *Desmognathus* in Virginia, **Organ** (2).

Territoriality.—Of *Hyla faber*, **Lutz**.

Venoms.—Poisonous function of *Rana chalconota* granular patches, **Liem**.

Voice.—Nightly records of calling activity in mixed anuran population in Texas over 4-year period, **Blair, W. F.** (2); Mating call recordings of *Hyla bivoca* sp. n., **Duellman & Hoyt**; Audiospectrograph of *Hyla rufigula* mating call, **Fouquette**; Calls of *Cyclorana platycephalus*, *Limnodynastes spenceri*, *Neobatrachus outor*, *Hyla rubella* and *Cyclorana cultripes*, **Lindgren & Main**; Mating call discrimination by *Pseudacris clarki* ♀, **Littlejohn** (1); Call structure of *Crinia* ♂, **Littlejohn** (3); Mechanics of sound production in North American *Bufo*, **McAlister** (2); Vocalization as an isolating mechanism in frogs **Martof** (1); Role of anuran mating calls in sexual isolation, **Mecham** (3); Voices of New South Wales frogs, **Moore, J. A.** (2); Call of *Kassina senegalensis*, **Wager** (2); Call of ♂ *Rana wagneri*, **Wager** (3).

GEOGRAPHICAL DISTRIBUTION

General: Zoogeography.—Zoogeographical study of the herpetofauna of Bulgaria, **Beškov**; Ranges of the amphibians of Florida, **Carr & Goin**; Herpetology of Argentina, **Cei** (1); Amphibian provinces of Uruguay, **Batabaroff**; Amphibians of North Carolina, **De Poe, Funderburg & Quay**; Geographic ranges of *Ambystoma macrodactylum* races in western North America, **Ferguson** (1); Zoogeographic affinities of the herpetofauna of Tishomingo Co., Mississippi, **Ferguson** (2); Zoogeographical study of Surinam, **Geijskes**; Distribution of *Ptychocheilus oxyrinchus* and *P. abyssinica* in Africa, **Guidé & Lamotte** (1); Zoogeographic relationships of the

herpetofauna of the Howard College Natural Area, Alabama, Holman (1); Amphibia of Canada and Alaska, a check list, Logier & Toner; Relationships of animal geography between Australia and other continents, Mertens (1); Preservation of animal provinces, Mertens (2); Australian frogs, distributional patterns and zoogeographic regions, Moore, J. A. (2); Of Amphibia in Southern Africa, Poynton (1); Biogeography of south-east Africa, Poynton (2); General anuran zoogeography, Reig (3); Statistical zoogeography, study of regional populations, Stugren & Rădulescu; Fossil and recent amphibians, Terent'ev (2); Distribution of *Gastrophryne mazatlanensis* in Northwestern Mexico and Southwestern U.S.A., Wake.

PALAEARCTIC REGION

Great Britain.—†Worcestershire, Upton Warren (*Rana temporaria*), Coope, Shotton & Strachan; Ireland (smooth newt), Fenton; Ireland, Ulster (*Triturus vulgaris*), Forsyth; Dorset (*Rana temporaria*, *Bufo bufo*), Hawthorne; †(Labyrinthodonts), Panchen & Walker.

Afghanistan.—(General), Leviton & Anderson.

Austria.—(Catalogue of Amphibia), Eiselt; Steiermark (*Pelobates fuscus*, *Bufo bufo*, *B. viridis*), Kepka in Kepka & Schuster.

Azores.—(*Rana esculenta*), Ulfstrand.

Belgium.—Hautes Fagnes (*Rana arvalis* and *Alytes obstetricans*), Meeuwen; Vervétois (key to species), Rose, L.

Black Sea area.—(Statistical survey), Stugren & Rădulescu.

Bulgaria.—(Herpetofauna), Beškov.

China.—†(Pliocene, *Rana*), Liu.

Corfu.—(General), Mertens (3).

Czechoslovakia.—(*Bombina bombina* and *B. variegata*), Lác (1); Orava River basin (general), Lác (2).

France.—Hautes-Pyrénées, Nèouville Mt., Estibère Lakes (*Rana temporaria*, *Triturus helveticus*), Angelier; Grande-Brière (general), Bodin; †Chilleux-aux-Bois (Burdigalian sands (?), *Salamandra*, *Rana*, *Bufo*), Chaline; Provence (general), Knoepffler (1).

Germany.—Dresden (*Rana dalmatina*), Fritzsche & Obst; R. Elbe lowlands (general), Garms; Gera (general), Kuhn (1); †(Fauna of the Solnhofener shales), Kuhn (2).

Greenland.—†(Devonian, *Ichthyostegalia*), Jarvik.

Italy.—Tuscany, Badizzano and Ritomboli caves (*Hydromantes italicus*), Lanza (1); Tuscany (cavernicole Anura and Urodela), Lanza (3); Tana del Leccio (*Hydromantes*), Lanza & Marcucci; Dolomites, Sesto (*Salamandra atra*), Marcuzzi.

Japan.—Okayama (*Rana nigromaculata nigromaculata* × *R. n. brevipoda*), Moriya.

Lebanon.—†Ksar' Akil (palaeolithic, *Hyla arborea*), Hooijer.

Mediterranean Region.—*Corsica* (general), Goux (1); *Corsica* (*Euproctus montanus*), Goux (2); (*Discoglossus* and others), Knoepffler (3).

Mongolia.—†(General), Orlov.

Netherlands.—Massane Reserve (general), Bree; Limburg (*Salamandra salamandra*), Fonteyne (1); Limburg (*Bombina variegata*), Fonteyne (2); (General), Wijngaarden.

North Africa.—Morocco (general notes), Bons; †Sahara (teeth replacement in *Capitosauria*), Coudron; †Morocco, Beni Mellal (Miocene, general), Hecht, Hoffstetter & Vergnaud; Morocco (unidentified tadpoles, *Alytes* or *Bombina* ?), Pasteur.

Poland.—Kraków-Chrzanów Ridge and R. Wisła valley (*Bombina*), Michałowski (1); †(Salientia and Caudata), Młynarski.

Portugal.—(General), Almaça.

Roumania.—(*Triturus cristatus*), Fuhr & Freytag; Sibbenbürgen (*Rana arvalis*), Stugren & Kohl; (General notes), Stugren & Popovici (1); (*Bombina* variation), Stugren & Popovici (2); *Circic-lasy* (*Rana ridibunda*), Vancea, Mindou & Simionescu.

Russia.—†Priural, Krasnyi Kamen (*Stegocephalia*), Chalyshev; †N. Dvina river (*Tupilakosaurus*), Shishkin.

Spain.—Guadalquivir (general), Valverde.

Yugoslavia.—Montenegro (neotenus *Triturus alpestris*, two new subsp.), Radovanović.

ORIENTAL REGION

Hong Kong.—(*Megophrys brachykolos* sp. n.), Inger & Romer.

India.—(Caecilians), Taylor; Portuguese India, Goa (*Rana tigrina*), Manacas.

Philippine Islands.—Cuernos de Negros (amphibian population), Brown & Alcalá.

AUSTRALIAN REGION AND POLYNESIA

Australia.—Jigalong, W. Australia, Lindgren & Main; S.W. Australia (*Crinia*), Littlejohn (3); †Canning Basin (Triassic Blina shale, general remains), McKenzie; Rottnest Island (*Crinia insignifera* populations), Main; Eastern New South Wales (review of Anura), Moore, J. A. (2).

New Zealand.—(Frogs), Barwick; (Introduced frogs), McCann; (*Leiopelma* spp.), Stephenson, E. M.

ETHIOPIAN REGION

Africa.—Kagera and Albert National Parks (general), Curry-Lindahl (2).

Central Africa.—*S. Rhodesia, Mt. Selinda* (*Arthroleptis xenodactyloides*), **Bruggen**; *Congo, Mayombe* (general), **Laurent** (2); *Congo, Stanley Pool* (Anura), **Skelton-Bourgeois** (2); *Tchad* (herpetofauna), **Wake & Kluge**.

East Africa.—(Systematic list of Batrachia), **Skelton-Bourgeois** (1).

West Africa.—*Spanish Guinea* (*Rana goliath*), **Cuspinera**; *French West Africa* (*Phrynobatrachus guineensis* and *P. alticola* spp. nov.), **Guibé & Lamotte** (2); *Sierra Leone, Tingi Hills* (*Bufo cristigilans* sp. n.), **Inger & Menzies**; (General), **Lamotte, Dzieduszycka & Lauwarier**; (General), **Lamotte & Perret** (2); *French Guinea, Mt. Nimba* (*Nectophrynoides occidentalis*), **Lamotte & Roy** (2).

Madagascar.—(Batrachia), **Guibé**; †(Trias, Stegocephalia), **Lehman**.

NEARCTIC REGION

Alaska.—(Check list), **Logier & Toner**.

Canada.—*Quebec Prov.* (general), **Alexandre**; *New Brunswick* (*Plethodon cinereus*), **Cook & Bleakney**; *Quebec province, Ibterville Co., Mt. Johnson* (*Hemidactylum acutatum*), **Denman**; *British Columbia, Storm Creek* (*Asaphus truei*), **Grant, J.**; (Check list), **Logier & Toner**; *Lake Manitoba* (herpetofauna), **Tamsitt**.

United States.—*Eastern United States* (*Desmognathus cookei*), **Valentine**.

North-Eastern States.—*Ohio* (County records), **Adler**; *Ohio* (new records), **Adler & Dennis**; *Eastern New York* (records), **Benton & Smiley**; *Ohio, Hamilton Co.* (herpetological records), **Collins & Haggard**; *West Virginia* (herpetology collection), **Green**; *Ohio* (general herpetology), **Hirschfeld & Collins**; *S.W. Virginia, Balsam Mts.* (general), **Organ** (2); *Central Indiana* (*Plethodon* spp.), **Reynolds**; *Illinois* (critical review, distribution, ecology, variation), **Smith, P. W.**; *Cattaraugus Co., Allegany Indian Reservation* (general), **Stewart**; *Illinois* †(list and bibliography of amphibians), **Techter**; *Michigan, Saddle Lake* (*Siren intermedia nettingi*), **Williams, J. E.**

South-Eastern States.—*Tennessee* (cave dwellers), **Barr**; *North Carolina* (*Siren i. intermedia*), **Collette & Gehlbach**; *North Carolina* (general list), **DePoe, Funderburg & Quay**; *Mississippi, Tishomingo Co.* (herpetofauna), **Ferguson** (2); *Alabama, Butler Co.* (*Phaeognathus hubrichti* gen. & sp. n.), **Highton** (2); *Alabama, Birmingham region, Howard College Natural Area* (general), **Holman** (1); †*Florida, Gilchrist Co.* (Miocene, *Proacris* gen. nov.), **Holman** (2); *S.E. Florida* (new populations of West Indian amphibians), **King**; *Alabama, Macon Co.* (*Hyla versicolor* × *H. avivoca*), **Mecham** (2); *North Carolina, Hyde and Tyrrell Counties* (general list), **Palmer & Whitehead**; *Louisiana* (*Acris crepitans* varieties), **Pyburn** (1); *Southern Appalachians* (*Plethodon* spp.), **Reynolds**; *Arkansas* (*Desmognathus fuscus* races), **Smith, C. C.**

Middle and Western States.—†*Wyoming, Niobrara Co.* (*Opisthotriton* gen. nov.), **Auffenberg**; *Texas, Edwards plateau* (*Eurycea*), **Baker, J. K.**; *Nevada,*

Clark Co., Colorado R. (*Hyla regilla*), **Banta** (2); *Oklahoma* (*Hyla avivoca*), **Blair & Lindsay**; *Minnesota* (*Bufo hemiophrys*), **Breckenridge & Tester**; *Texas* (*Acris crepitans* varieties), **Pyburn** (1); *Texas* (collection), **Webb & Packard**.

Pacific Coast States.—*California* (salamanders), **Brame, A. (1)**; *Oregon* (*Taricha granulosa*), **Livesey & Wyllie**; *Sonoma County, Pepperwood Creek* (*Taricha*), **Twitty** (1).

CENTRAL AMERICA

Central America.—(*Rhinophrynus dorsalis*), **Nelson & Hoyt**.

British Honduras.—(Herpetofauna), **Neill & Allen**.

Canal Zone.—(*Hyla ruftela* sp. n.), **Fouquette**; *Panama, Chiriquí Prov., Volcán Baru* (*Magnadigitia marmorea* sp. nov.), **Tanner & Brame**.

Costa Rica.—(Salamanders), **Brame, A. (2)**; (*Crepidius*), **Savage & Kluge**.

Guatemala.—(*Bufo valliceps* races), **Baylor & Stuart**.

Mexico.—(*Ambystoma rosaceum*), **Anderson, J. D. (2)**; *Volcán Parícutin* (effect on amphibians), **Burt**; (New records), **Chrapliwy, Williams & Smith**; (*Ptychohyla* spp. nov.), **Duellman** (1); *Chiapas* (*Hyla chaneque* sp. n.), **Duellman** (2); *Oaxaca, Vista Hermosa* (general), **Duellman** (3); *Chiapas* (*Hyla bivocata* sp. n.), **Duellman & Hoyt**; *Isla Cerralvo* (*Scaphiopus couchi*), **Etheridge**; *Sinaloa, El Dorado* (general), **Fugler & Dixon**.

SOUTH AMERICA

Andes.—*Of Peru and Bolivia* (frogs), **Vellard**.

Argentina.—†*Chubut province, Laguna del Hunco* (Eocene, *Shelania pascuali* gen. & sp. nov.), **Casamiquela** (1); †*Patagonia* (*Notobatrachus*), **Casamiquela** (2); (General), **Cei** (1); *Misiones* (Anura), **Gallardo** (1); *Buenos Aires* (*Hyla raddiana*), **Gallardo** (4); (Herpetology), **Gallardo** (6); †*Salta* prov. (*Saltenia* gen. nov.), **Reig** (1); †*Patagonia* (Jurassic, Anura), **Reig** (2).

Bolivia.—(*Bufo* spp. n.), **Gallardo** (5).

Brazil.—(*Bufo manicorensis* sp. n.), **Gallardo** (5); (*Habrahyla eiselti* gen. & sp. nov.), **Goin** (3).

Chile.—(*Bufo spinulosus* and *B. arunco*), **Cei** (5); (*Eupsophus*), **Grandison** (1).

Colombia.—(*Amblyophrynus ingeri* gen. & sp. nov.), **Cochran & Goin**; †(General), **Porta**.

Ecuador.—(*Cochranella* spp. n.), **Goin** (2).

Surinam.—(General), **Audretsch** (1) & (2); (General), **Geijskes**.

Uruguay.—(General), **Chebataroff**; (General), **Klappenbach**.

Venezuela.—(*Salientia*), **Rivero**.

PALAEONTOLOGY AND GEOLOGICAL DISTRIBUTION

Carboniferous.—(Labyrinthodonts), **Panchen & Walker.**

Cretaceous.—(Wyoming, *Opisthotriton* gen. nov.), **Auffenberg**; (*Saltentia* gen. nov.), **Reig** (1).

Devonian.—(*Ichthyostegalia*), **Jarvik.**

Jurassic.—(*Notobatrachus*), **Casamiquela** (1); (Patagonia, Anura), **Reig** (2).

Miocene.—*Burdigalian* (?) (Chilleurs-aux-Bois, France, *Salamandra*, *Rana*, *Bufo*), **Chaline**; (Morocco, general), **Hecht, Hoffstetter & Vergnaud**; (Florida, *Proacris* gen. nov.), **Holman** (2).

Permian.—"Kupferschiefer" (general), **Kuhn** (1).

Pleistocene.—(Worcestershire, *Rana temporaria*), **Coope, Shotton & Strachan**; (Australia, speciation of *Urinia*), **Littlejohn** (3).

Pliocene.—(China, *Rana*), **Liu**; (Poland, Salientia and Caudata), **Mlynarski.**

Quaternary.—*Palaeolithic* (Lebanon, *Hyla arborea*), **Hootjer.**

Tertiary.—*Eotertiary* (Chubut province, Argentina, *Shelania pascuali* gen. & sp. nov.), **Casamiquela** (1).

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THE ORDERS, AND FAMILIES AND GENERA WITHIN THEM, ARE ALPHABETICALLY ARRANGED.

References to the "Titles" are by the name(s) of the Author(s) printed in clarendon type.

Note.—The arrangement here used follows in general that of Romer's "Vertebrate Paleontology".

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†*Discoglossus* sp., maxillare, ilium, urostyles and limb bones, Pliocene, Weze, Poland, **Młynarski**.

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†*Hyla arborea*, from a Palaeolithic rock shelter in Lebanon, **Hooijer**.

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†Pipidae from the Miocene of Beni Mellal, Morocco, Hecht, Hoffstetter & Vergnaud.

Hymenochirus boettgeri, embryonic development, evolution and homologies of sensory lines and dermal bones of the cheek, Stensiö; *H. boulengeri*, general notes, aquarium requirements, breeding, photos., Haas; Breeding in captivity, photos., Yuhas.

Pipa parva, from Venezuela, Maracaibo basin and Falcón region; N.E. Colombia, Rivero; *P. pipa*, photos. 1 col., Cochran (2); Reproductive habits, brooding care, egg development and emergence of young, photos., Rabb; Eastern Venezuela, Trinidad and the Guianas to Matto Grosso; Colombia to Peru and probably Bolivia, Rivero.

†*Saltenia* gen. nov. p. 4 of *Aglossa* (?) genotype *S. ibañezii* sp. nov. p. 4, pl. 1, figs. 1, 2; Alemania (Dto. de Viñas, Salta prov.), N.W. Argentina, Lower Cretaceous, Reig (1).

†*Shelania* gen. nov. p. 17 of Pipidae, type-species *S. shalensis* sp. nov. p. 18, fig. 1; Eocene, Laguna del Hunco, Chubut province, Argentina, Casamiquela (1).

Xenopus, the pineal gland and body lightening in larvae, Bagnara (1); Larvae, chemically limited retardation of growth, in spite of the influence of acid phosphatase, in tail regeneration, Hahn, Niehus, Scholl & Lehmann; Thyrostatic effect of $KClO_4$, Pfugfelder; Effect of 5-fluorodeoxyuridine on embryos, Tencer (1); Electron microscopical observations on larval tissues, Weber; *X. fraseri*, from Mayombe, Congo. Distinguished from *X. tropicalis*, Laurent (2); *X. laevis*, onset of pineal and hypophyseal regulation of melanophores, Bagnara (2); Ultrastructural mechanisms of gastrulation and neurulation, Balinsky, B. I.; Mode of excretion of ammonia and urea, Balinsky & Baldwin; Transfer of primordial germ-cells, Blackler & Fischberg; Morphogenetic effects of lipoic acid on embryos, Brachet; Ageing, Brocas & Verzar; Photos., Cochran (2); Existence of temporal specificity in reaggregation of mid-gastrula cells, Curtis; Effect of ATP on late gastrulae explants, Deuchar (1); Increased ^{32}P exchange between pyrophosphate and adenosine triphosphate in the presence of added L-leucine in embryos, Deuchar (2); Relationship with *Marsupiodella africana* in the Kromboom River, Cape Province, S. Africa, Dick; Sources and nature of salivary secretions in adult, Francis; Lens regeneration from cornea, Freeman & Overton; Egg involucra of *Discoglossus pictus*, Ghiara; Response of melanophores to catecholamines, Graham; Form and function of larval fore-gut with reference to the *manicotto glandulare*, Griffiths; Treatment with 4-methyl-2-thiourea causing intense degeneration of the islets of Langerhans, Guardabassi; Nuclear transplantation between 2 subspecies, Gurdon; Differential behaviour, Haubrich (1); Hierarchical behaviour, Haubrich (2); Changes in serum proteins during metamorphosis, Herner & Frieden (1); Cell degeneration in ventral horn of larva, Hughes, A.; Effect of limb ablation on neurones, Hughes & Lewis; Dominant foods, seasonal and altitudinal variation in diet and species-prey size relations in the Congo, Inger & Marx; Development of regeneration blastemas implanted into the brain, Jordan; Origin, importance and fate of melanocytes in cerebral fluid, Komnick; Effect of food on speed of larval development, Kucias; Regeneration of transected nervous connections in the brain, Kwiatkowski; Bile pigment excretions, Lester & Schmid; Effects of axon severance in cholinergic neurones of larva, Lewis & Hughes; Development of peripheral nerves, Peters, A.; Blood vessels of developing spinal cord, Sims (1); Treatment with phenylthiourea, Sims (2); Effect of brain injuries on development, Srebro; Experiment to discover whether artificially induced "desiccation" affected subcommissural organ, Steyn; Effect of morphogenetic inhibitor on water balance of embryo, Tuft (1); Role of H_2O -regulating mechanisms in morphogenesis, Tuft (2); Egg laying habitat, Van Dijk; Alterations in connective tissue and intestine produced by hypervitaminosis A, Weissmann (2); Importance of ilio-sacral joint in locomotion, Whiting; Modification of Benz's method

for histochemical localization of cathepsins in tail, **Wróblewski**; Cathoptic activity in the tail during development, metamorphosis and regeneration, **Wróblewski & Gzybek**; *X. tropicalis*, from Mayombe, Congo. Distinguished from *X. fraseri*, **Laurent** (2).

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Key to Venezuelan genera, **Rivero**.

Lysapsus limellus, diagnostic features of subspecies tabled, *L. limellus bolivianus* **subsp. nov.** p. 127; **Reyes**, Bolivia, *L. l. laevis*, from Brazil, description, colour, dimensions and distribution, and *L. l. limellus* from Paraguay, *L. mantidactylus* (Cope) from Uruguay, external and skeletal characters, dimensions and distribution; synonymy includes *Pseudis mantidactyla* Cope 1862, **Gallardo** (8).

Pseudis mantidactyla Cope, see *Lysapsus mantidactylus* (Cope), **Gallardo** (8); *P. paradoxa* Surinam, photo., **Audretsch** (2); Photos., **Cochran** (2); *P. paradoxus*, external morphology, skeleton, diagnostic features and interrelationships of subspecies, figs., **Gallardo** (8); Range, habits and locality records, Venezuela, **Rivero**; *P. paradoxa*, chromosome number, **Saez-N. Brum**; *P. paradoxus bolodactylus*, from Brazil, description, dimensions, variation and distribution, *P. p. caribensis* **subsp. nov.** p. 116, fig. 4; Mayaro Bay, Trinidad, B.W.I., *P. p. fuscus*, from Brazil, description, dimensions and distribution, *P. p. occidentalis* **subsp. nov.** p. 122, fig. 4; El Pailón, Bolivia, *P. p. paradoxus*, from British Guiana, description, dimensions and distribution, *P. p. platensis* **subsp. nov.** p. 120, fig. 4; Colonia Nueva Italia, Dept. Villeta, Paraguay, **Gallardo** (8).

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Form and function of larval fore-gut with reference to the *manicoto glandulare*, **Griffiths**; Key to Venezuelan genera, **Rivero**.

†Ranidae from the Miocene of Beni Mellal, Morocco, **Hecht, Hoffstetter & Vergnaud**.

Abroscaphus **Laurent**, replaced in synonymy of *Arthroleptis* A. Smith, **Laurent** (1).

Anhydrophryne rattayii **Hewitt**, taxonomic status, osteological characters, **Laurent** (1).

Arthroleptella **Hewitt**, taxonomic status of genera, osteological analysis and phylogeny of *laurenci* Loveridge 1954, *hewitti* **hewitti** Fitz Simons 1947, and *h. minor* Fitz Simons 1947, **Laurent** (1).

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Arthroleptis adolfi-friederici leleupi **subsp. nov.** p. 323, Bagilo, Uluguru Mts., Tanganyika Territory, **Skelton-Bourgeois** (1); *A. stenodactylus*, dominant foods, seasonal and altitudinal variation in diet and species-prey size relations in the Congo, **Inger & Marx**; *A. wahlbergi* A. Smith 1849, key to taxonomic status of genus and included species, **Laurent** (1); *A. zenodactyloides*, from Chirinda forest, S. Rhodesia, **Bruggen**.

Astylosterninae, of Mayombe, Congo. Key to *T. robustus*, *A. diadematus* and *S. gabonicus* (q.v.), **Laurent** (2).

Astylosternus diadematus, first record from Congo; comparison with Cameroon specimens, note on variation in proportions, **Laurent** (2).

Aubria subsigillata, from Stanley Pool, Congo; tadpoles, **Skelton-Bourgeois** (2).

Cacosterninae, considered diphyletic, phylogenetic study of genera, **Laurent** (1).

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Conraua robusta, generic type; note on validity of specific name, **Perret**.

Dendrobates, references at present retained under DENDROBATIDAE.

Dicroglossus occipitalis, from Stanley Pool, Congo; dental formula, measurements, coloration, **Skelton-Bourgeois** (2).

Hemius marmoratus, photo., **Cochran** (2); Photo., ecology in Albert and Kagera National Parks, Africa, **Curry-Lindahl** (2); From Tchad, Africa, **Wake & Kluge**.

Hylarana acutirostris, specific name preoccupied in *Rana*; replaced by *parkeriana* **Mertens**. Cameroon race becomes *H. parkeriana longipes* **Perret**, **Perret**; *H. albolabris*, from Stanley Pool, Congo; dental formula, measurements, coloration, **Skelton-Bourgeois** (2).

Phrynobatrachinae, osteological comparison with Arthroleptinae, phylogenetic relationships and taxonomy, **Laurent** (1).

Phrynobatrachus alticola **sp. nov.** p. 574, figs. 3 & 4; Mt. Nimba region, Guinea, W. Africa, **Guibé & Lamotte** (2); *P. anotis* and *P. eryptotis*, dominant foods, seasonal and altitudinal variation in diet and species-prey size relations in the Congo, **Inger & Marx**; *P. guineensis* **sp. nov.** p. 571, figs. 1 & 2; Mont Tonkoui, Ivory Coast, **Guibé & Lamotte** (2); *P. gutturosus*, dominant foods, seasonal and altitudinal variation in diet and species-prey size relations in the Congo, **Inger & Marx**; *P. natalensis*, ultrastructural mechanisms of gastrulation and neurulation, **Balinsky, B. I.**; Photo., **Cochran** (2); *P. natalensis*, *P. parvulus* and *P. perpalmaris*, dominant foods, seasonal and altitudinal variation in diet and species-prey size relations in the Congo, **Inger & Marx**.

Phyllobates, references at present retained under DENDROBATIDAE.

Platymantis boulengeri, ool. photo., **Cochran** (2).

Prostherapis, references at present retained under DENDROBATIDAE.

Ptychadena abyssinica **Peters** 1881, Africa, description, biometrics, distribution, comparison with *P. oxyrhynchus*; synonymy includes *P. gondokorensis* **Werner** 1907, *P. aberae* **Ahl** 1923, *P. oxyrhynchus oxyrhynchus* **Lov.** and *P. o. migiurtina* **Scort.** 1933,

Guibé & Lamotte (1); *P. macarthysensis*, W. Africa, larval structure, development and metamorphosis, biometrics, figs., Lamotte & Perret (1); From Tchad, Africa, morphology, Wake & Kluge; *P. mascareniensis*, larval structure, development and metamorphosis, biometrics, Camerouns and Egypt, Lamotte & Perret (1); *P. m. mascareniensis*, photo., ecology in Albert and Kagera National Parks, Africa, Curry-Lindahl (2); *P. oxyrhynchus* Smith 1849, Africa, description, biometrics, distribution, comparison with *P. abyssinica*; synonymy includes *P. gribinguiensis* Angel, and *P. oxyrhynchus gribinguiensis* Loveridge, Guibé & Lamotte (1); *P. oxyrhyncha*, from Tchad, Africa, morphology, Wake & Kluge; *P. perreti*, larval structure, development and metamorphosis, biometrics, W. Africa, Lamotte & Perret (1); *P. submascareniensis*, larval development and metamorphosis desc'd. and fig'd.; phalangeal and dental formulae. Diagnosis of adult; vomerine teeth absent, Lamotte, Dzieduszycka & Lauwarier; *P. tournieri*, diagnosis of species, fig. Larval development and metamorphosis desc'd., fig'd. Phalangeal formula; dental formula, Lamotte, Dzieduszycka & Lauwarier; *P. trinodis*, adult and larva desc'd., fig'd. Larval development and metamorphosis; note on dental formula, Lamotte, Dzieduszycka & Lauwarier; From Tchad, Africa, morphology, Wake & Kluge; See also references under *Rana*.

Rana agilis, from swamps between Pierre-Fendue and Bréca, Bodin.

Rana areolata areolata, from E. Texas, Webb & Packard; *R. a. circulosa*, variation, distribution and ecology in Illinois, photo., Smith, P. W.

Rana arvalis, from R. Elbe lowlands, Germany, Garms; Hautes Fagnes, Belgium, Meeuwen; Morphogenetic study of cerebellar nuclei, Rüdeberg; Induction of ovulation out of breeding season, Štefanová & Romanovský; Larvae fed on snails in aquarium, effect on development, Stolk (11); Kétyi Nyir, Siebenbürgen, biology, Stugren & Kohl; Statistical survey, Black Sea area, Stugren & Rădulescu; Actively acquired tolerance to skin heterografts, Vyazov & Sorokina; *R. a. wolterstorffi* Szászregén, Siebenbürgen, biology, Stugren & Kohl.

Rana aurora aurora and *R. a. cascadeae* shown, on the basis of experimental crosses, to be reproductively isolated and incapable of interbreeding; concluded that the two are separate species and should be designated as *Rana aurora* Baird & Girard, and *R. cascadeae* Slater, Porter.

Rana blythi, photo., Cochran (2).

Rana boylei, histology of pancreatic islet, Miller.

Rana cancrivora, osmotic and ionic regulation studies, Gordon, Schmidt-Nielsen & Kelly.

Rana capito, photo., Florida, Carr & Goin.

Rana cascadeae Slater, shown to be a distinct species on the basis of experimental crosses with *R. aurora*, Porter.

Rana catebeiana, calling and spawning seasons in Texas, Blair, W. F. (2); Photo., Florida, Carr & Goin; Photos., Cochran (2); Effect of sodium

fluoroacetate on blood glucose levels, Cole; Extremity vein endothelial nuclei, Donahue, Connor & Manner; Relationship of O₂ consumption to body weight and starvation, Fanslow; Comparative activity of thyroxine and its analogues in tadpoles, Frieden & Westmark; Biomechanics of jumping at prey, photos., Gans (4); Regeneration and stability of lateral line organs, Helff; Changes in serum proteins during metamorphosis, Herner & Frieden (1); Histocompatibility genetics of isolated populations, Hildemann & Haas; Host to a new *Ophiotaenia*, Jones, Cheng and Gillespie; Release of Ca-45 from nerves during electrical activity, Koketsu & Miyamoto (1); Bile pigment excretions in larvae, Lester & Schmid; Effect of chemicals on tadpole respiration, Lewis & Frieden; Isolated retina used to show mechanism of light adaptation, Lipetz; Construction of ileostomies and colostomies in adult, Massaro; Localization of acid and alkaline phosphatase in pituitary, Ortman & Fallon; Effect of thyroxine on protein synthesis in the tadpole, Paik & Cohen; Influence of dietary factors and hormones on respiratory metabolism in larvae and adults, Pryor & Tipton; Cover photo., climbing an electric fence, Schmid, F. C.; Oxyntic cells in the gastric glands, Sedar; Embryology of larval corium, Shimozawa; Bodies of Eberth in tadpole skin, Singer & Salpeter; Variation, distribution and ecology in Illinois, photo., Smith, P. W.; Influence of parathyroids on bone, Talmage & Yoshida; Action of oxytoxin and temperature on glomerular filtrate, Uranga; Total blood volume determination by radioisotope methods, Wilson, Hansard & Cole; Preparation of monolayer cell cultures from various tissues, Wolf, Quimby, Pyle & Dexter; Blood sugar studies, Wright, P. A.

Rana chalconota Schlegel, taxonomic status, comparison with *R. labialis* Schlegel; measurements given; evidence to support Boulenger's view (1920) that *labialis* is not a species distinct from *chalconota*; note on function of poisonous granular patches, Liem.

Rana chensinensis, postembryonic multiplication of muscle fibres, Lobytsev.

Rana christyi, photo., Cochran (2).

Rana clamata, embryonic development, evolution and homologies of sensory lines and dermal bones of the cheek, Stensiö.

Rana clamitans, photo., Florida, Carr & Goin; Photo., Cochran (2); Dimensionless characteristics of duration of development, Dettlaff, T. A. & A. A. (1) & (2); Comparative activity of thyroxine and its analogues in tadpoles, Frieden & Westmark; Absorption of amino acids in the intestine, Gagnon; Changes in serum proteins during metamorphosis, Herner & Frieden (1); Construction of ileostomies and colostomies in adult, Massaro; Bodies of Eberth in tadpole skin, Singer & Salpeter; Caught in mousetraps near Ithaca, New York; stomach contents and habitats, Whitaker; *R. c. melanota*, variation, distribution and ecology in Illinois, photo., Smith, P. W.

Rana curtipes, metamorphosis observations, Lobo.

Rana cyanophlyctis, action of mammalian hormones on hypophysectomized specimens, Ramaswami & Lakshman.

Rana dalmatina, morphogenesis of the digestive apparatus, **Albert & Cambar**; Origin of formative material for ♀ genital region, **Dechambe**; Different reactions of pronephros to thyroid hormone, **Dechambe & Cambar**; New record in C. Europe, **Fritzsche & Obst**; ♂ sexual cycle, **Lanza** (2); In Corfu, synonymy, identification and distribution, **Mertens** (3); Induction of ovulation out of breeding season, **Stefanová & Romanovský**; Statistical survey, Black Sea area, **Stugren & Rădulescu**.

Rana esculenta, in Portugal, distribution, systematics and ecology, **Almaça**; Effect of skin secretions on muscle, **Bartha, Faiszt & Jendrassik**; From the Grande-Brière region, **Bodin**; Spinal cord potential generation by cutaneous nerve stimulation, **Bravo & Fernandez de Molina**; Histology of the optic tectum, **Capanna** (1); Ultrastructure of resting and contracted striated muscle, **Carlsen, Knappeis & Buchthal**; Action of intermediin on thyroid function, **Čehović**; Photos., **Cochran** (2); Effect of penicillin and streptomycin on growth and development of tadpoles, **Dąbrowski**; Physiological and biochemical properties of dehydrated muscles and heart, **Dydyńska**; Electrophysiology of muscle fibres, **Eitzensperger** (1) & (2); Growth of pronephric system quantitatively analysed, **Fox, H.** (1); Use of males in new test for early pregnancy diagnosis, **Frassinetti & Lanza**; Histology and histochemistry of macrothryocytes, **Gabe**; From R. Elbe lowlands, Germany, **Garms**; Egg involution cf. *Discoglossus pictus*, **Ghiara**; Role of semi-circular canals, **Gribenski**; Transplantation of *R. pipiens* nuclei into enucleated eggs, and their early development, **Hennen**; Habits in captivity and in natural habitats modified by man, **Heusser**; Effect of lysine-vasotoxin on oxytocin, competitive inhibition of antidiuresis, **Jard & Morel**; Effect of temperature on O₂ linkage in blood, **Krüger**; In the Orava River basin, Czechoslovakia, ecology, measurements, altitude limit, **Lác** (2); ♂ sexual cycle, **Lanza** (2); Yolk formation in oocytes, **Lanzavecchia**; Effect of powdered pineal gland on tadpole development, **Lawinski**; Influence of gibberellins on embryo, **Liotti** (1); Effect of chlorotetracycline on larval tail regeneration, **Manelli** (1); Effects of low temperatures on eggs and embryos, **Manelli** (2); Stages in development, figs., **Manelli & Margaritora**; Effect of loss of blastomere material on development of egg, **Milaire**; Neurohypophyseal hormones, **Morel, Maetz, Acher et al.**; Morphogenetic functions of nucleus during early development, **Neifakh**; Influence of Ca on K-contracture of "slow" and "fast" skeletal muscle fibres, **Pauschinger & Brecht**; Influence of the atmosphere enriched in CO₂ and O₂ on skin capillaries, **Pecozeko**; Isolation of steroids with phenolic and ketonic fractions from sexually active frogs, **Raunich** (1); Morphogenetic potential of the lateral mesoderm in tail bud stages, **Raunich** (3); Neural crest transplantation from neurulae to belly wall, **Raunich** (4); Differentiation of explanted cephalic neural folds from neurulae, **Raunich** (5); Development of neural fold material after homoplastic transplantation into the orbit of differentiated tadpoles, **Raunich** (6); Presence of p-hydroxy-benzylacetone in extracts, **Raunich** (7); Effect of testosterone on early embryology, **Rodriguez-López** (1); Effect of 2-4-dichlorophenoxyacetic acid and its Na salt on the motility of spermatozoa, **Rodriguez-López** (2); Study of nervous system development by transplantation experiments, **Rossi** (3); Nervous regulation during parabiosis, **Rossi** (4) & (5); Survival and fecundity of spermatozoa in *in vitro*

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Rana frontalis, dominant foods, seasonal and altitudinal variation in diet and species-prey size relations in the Congo, **Inger & Marx**.

Rana fuscipula, photo., **Cochran** (2); Dominant foods, seasonal and altitudinal variation in diet and species-prey size relations in the Congo, **Inger & Marx**; *R. f. nutti*, photo., ecology in Kagera and Albert National Parks, Africa, **Curry-Lindahl** (2).

Rana goliath, from Spanish Guinea, photo., **Cuspinera**.

Rana graeca, photo., **Cochran** (2); Statistical survey, Black Sea area, **Stugren & Rădulescu**.

Rana grandisonae, dominant foods, seasonal and altitudinal variation in diet and species-prey size relations in the Congo, **Inger & Marx**.

Rana grisea, eaten in New Guinea, **Tyler** (2).

Rana grylio, photo., Florida, **Carr & Goin**; Enzyme changes and nucleic acid metabolism during metamorphosis, **Finamore & Frieden**; Comparative activity of thyroxine and its analogues in tadpoles, **Frieden & Westmark**; Changes in serum proteins during metamorphosis, **Herner & Frieden** (1); Changes in nature of red cell proteins during metamorphosis, **Herner & Frieden** (2); Effect of chemicals on tadpole respiration, **Lewis & Frieden**; Bodies of Eberth in tadpole skin, **Singer & Salpeter**.

Rana heckscheri, photo., Florida, **Carr & Goin**; Changes in serum proteins during metamorphosis, **Herner & Frieden** (1); Effect of chemicals on tadpole respiration, **Lewis & Frieden**.

Rana hexadactyla, haemoglobin content and sedimentation rate in blood, **Cherian, Vasu & Krishnaswamy**.

Rana hosii, col. photo., **Cochran** (2).

Rana iberica, in Portugal, distribution and ecology, **Almaça**.

Rana japonica, effect of Ca reduction on electrical activities of muscle membrane, **Hisada & Miyamoto**; Polarized light microscopy of single muscle fibre, **Washio, Kakiuchi & Tamasige**; Electrophysiological study on effects of anaesthetics on isolated muscle fibres, **Yamaguchi**; Induction of gonad masculinization by pituitary grafts in heat-treated hypophysectomized larvae, **Yoshikura**.

Rana labialis Schlegel, further evidence to support Boulenger's view (1920) that *labialis* is not a species distinct from *chalconota*, Liem.

Rana mascareniensis, dominant foods, seasonal and altitudinal variation in diet and species-prey size relations in the Congo, Inger & Marx.

Rana melanota, col. photo., Cochran (2).

Rana moeruensis, photo., Cochran (2).

Rana montezumae, from Jalisco, Mexico, Chrapliwy, Williams & Smith.

Rana nigromaculata, histology of seasonal changes of testis and thumb pad, Iwasawa & Asai; Histochemistry of kidney, Okada; Effects of metabolic inhibitors and narcotics on ^{32}P -outflux from retina, Oki & Honjo; Relationship between mitochondria and yolk platelets in embryonic cells, Sung; "On" and "off" responses in the lower olfactory pathway, Takagi & Shibuya; *R. n. brevipoda* and *R. n. nigromaculata*, nuclear transplantation experiments, Sambuichi; *R. n. nigromaculata*, topography of presumptive rudiments in endoderm of neurula, Tahara & Nakamura.

Rana occipitalis, from Tchad, Africa, Wake & Kluge.

Rana oxyrhyncha, photo., Cochran (2).

Rana palmipes, range, additional locality records, habits, Venezuela, Rivero.

Rana palustris, col. photo., Cochran (2); Dimensionless characteristics of duration of development, Dettlaff, T. A. & A. A. (1) & (2); Variation, distribution and ecology in Illinois, photo., Smith, F. W.

Rana perezi, photo., Cochran (2).

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